The Effects of *Eryngium campestre* Extracts on Glutathione-S-Transferase, Glutathione Peroxidase and Catalase Enzyme Activities

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Eryngium campestre L. (Apiaceae) are known in Turkish folk medicine as 'Bogadikeni' and wildly distributed in all parts of Turkey. Infusion of different parts of this species are well known as diuretic, appetizer and stimulant. In the present study, methanol extracts from flowers and leaves of E. campestre were evaluated for their potential medicinal value in terms of biological targets participate in antioxidant defense such as glutathione-S-transferase (GST), glutathione peroxidase (GPx) and catalase (CAT). Results obtained show that the methanol extract of flowers were found to have higher amount of phenolic and flavonoid contents than the leaves. The evaluation of extracts against biological target revealed that, the maximum GST enzymes inhibition activities was observed with methanol extract from flowers of E. campestre with IC₅₀ value of 363 ng/mL. In this study, none of the extracts evaluated were shown any reasonable GPx and CAT inhibition.

Key words: Eryngium campestre, Glutathione-S-transferase, Glutathione peroxidase, Catalase

Eryngium campestre Ekstrelerinin Glutatyon-S-Transferaz, Glutatyon Peroksidaz ve Katalaz Enzim Aktiviteleri Üzerine Etkileri

Geleneksel Türk tıbbında 'Boğa dikeni' olarak bilinen *Eryngium campestre* L. (Apiaceae), Türkiye'nin her bölgesinde genişçe yayılım göstermektedir. Bu bitkinin çeşitli kısımlarından hazırlanan infüzyonların, diüretik, iştah açıçı ve uyarıcı olarak etki gösterdiği bilinmektedir. Bu çalışmada, E. campestre'nin çiçek ve yapraklarından elde edilen metanol özütlerinin, antioksidan savunma sistemlerinde yer alan glutatyon-S-transferaz (GST), glutatyon peroksidaz (GPx) ve katalaz (CAT) enzimleri üzerine etkileri incelenmiştir. Elde edilen sonuçlarda, bitkinin çiçek kısımlarının metanol özütlerinin fenolik ve flavonoid içeriklerinin, yapraklarından elde edilen özütlerden daha yüksek değere sahip olduğu bulunmuştur. Biyolojik hedeflere karşı özütlerin etkilerinin değerlendirilmesinde ise, maksimum GST inhibisyonunun, *E. campestre*'nin çiçek kısımından elde edilen ekstrelerle sağlandığı ve IC₅₀ değerinin 363 ng/mL olduğu görülmüştür. Bu çalışmada değerlendirilen özütlerin GPx ve CAT inhibisyonu sağlamadığı görülmüştür.

Anahtar kelimeler: Eryngium campestre, Glutatyon-S-transferaz, Glutatyon peroksidaz, Katalaz

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INTRODUCTION

The genus Eryngium L. belonging to the subfamily Saniculoideae of the family of Apiaceae is represented by 317 taxa worldwide and are known to contain flavonoids, coumarins and triterpene saponins (1, 2). Ervngium campestre L. grows mainly in central and southern Europe, northern Africa and most of the Asia (3). In Turkish folk medicine E. campestre is known as "Bogadikeni" and wildly distributed in all parts of Turkey (4, 5). Infusion of aerial and root parts of this species are used in folk medicine to treat cough, urinary infections, disturbed functions of kidney, increased urine secretion and for regulation of the function of prostate (6). Also, the root of E. campestre is known as diuretic, appetizer, stimulant and aphrodisiac (1). Phytochemical studies show that the aerial parts and roots of *E. campestre* contain flavonoids, monoterpene glycosides, coumarin derivatives and saponins (7, 8).

The resistance of human tumor to multiple chemotherapeutic drugs has been recognized as a major reason for the failure of cancer therapy. Multidrug resistance can consist many mechanisms include alternations in drug transport, enhanced DNA repair, alternations in target proteins and alternations in drug metabolism such as; the detoxifying enzymes that are involved the glutathione dependent enzymes, glutathione-S-transferase (GST) and glutathione peroxidase (GPx) (9, 10). The glutathione-S-transferases (GSTs; EC: 2.1.5.18) are major phase II detoxification enzymes and most of the isoenzymes are located in the cytosol. They catalyze the conjugation of glutathione (GSH) to variety of exogenous and endogenous electrophilic compounds (11), serve roles development of drug resistance as inhibitor of the mitogen-activated protein (MAP) kinase pathway (12-14). Glutathione peroxidase (GPx; EC: 1.11.1.19) is the important enzyme which functions to protect the cell from oxidative damage by catalyzing the reduction hydroperoxides, including hvdrogen peroxide. Catalase (CAT; EC: 1.11.1.6) is a very important enzymes of living organisms which catalyzes the decomposition of hydrogen peroxide to water and oxygen. The

balance between activities and the intracellular levels of antioxidant enzymes are very important for the survival of organisms and their health. Previous studies have shown that, chemotherapy causes several side effects on biological system. During cancer therapy, some chemotherapeutic agents generate free radicals which cause apoptosis of target cells, but antioxidant enzymes may scavenge these radical molecules (15, 16) and may reduce the effectiveness of therapeutic agents. Thus, in this way, antioxidant enzymes inhibition may the treatment of proliferative improve diseases.

The aim of the present study is to evaluate the total amount of the phenolic and flavonoid contents of methanol extract obtained from flowers and leaves of *E. campestre* and to determine their inhibitory effect on the glutathione-S-transferase, glutathione peroxidase and catalase enzymes.

EXPERIMENTAL

Chemicals

In this study, 4-aminoantipyrine (4-AP), hydrogen peroxide (H₂O₂) and sodium azide (NaN₃) were purchased from Acros, USA. Ethylenediaminetetraacetic acid (EDTA), Folin-Ciocalteu reagent, reduced glutathione (GSH), glutathione reductase (GR), horseraddish peroxidase (HRP), catalase (CAT), gallic acid and quercetin hydrate were supplied from Sigma Aldrich, Germany. Nicotineamide adenine dinucleotide phosphate (NADPH) was purchased from Gerbu, Germany. All other chemicals used were analytical grade and purchased from Sigma Aldrich, Germany.

Plant material

Plant samples of *Eryngium campestre* L. were collected in July 2010 from Ankara-Turkey at their flowering season. The plant specimens were botanically identified by Associated Professor Dr. Fatmagül Geven, in Department of Biology, Ankara University. The plant specimens with their localities and the necessary field records were written and enumerated as voucher specimen number, FG 2010-18. Voucher specimens were deposited

in herbarium department at Ankara University.

Extraction of plant

Different parts of plant samples (flowers and leaves) were washed with tap water then dried at room temperature. For extraction, the previously reported procedure was used with some modifications (17). Basically, the plant samples were grounded with liquid nitrogen, to obtain fine powder of samples. Then, these samples were extracted with methanol for 24 hour at 4°C, with sample to solvent ratio of 1:10 (w/v). On the following day, the solvent was evaporated at 40°C until dryness, the obtained product dissolved in DMSO and kept at dark (4°C) until they are used.

Total polyphenol content

The total concentration of phenolic content of extracts were determined by employing the method described previously Slinkard& Singleton (18) with use of 2% (w/v) sodium carbonate solution and Folin-Ciocalteu's reagent (1:10 diluted with distilled water). The absorbance changes were spectrophotometrically measured as endpoints at 750 nm using multimode microplate reader (SpecraMax M2e, USA). The total polyphenol content (TPC) of extract was expressed as milligrams of gallic acid equivalent (GAE)/ L of plant extract (mg GAE/L) where the standard curve used were prepared with 0-250 mg/L of gallic acid (GA) in DMSO.

Total flavonoid content

The total concentration of flavonoids of extracts were determined by employing the aluminum chloride colorimetric method with use of 95 % ethanol, 10 % aluminum chloride, 1M sodium acetate and DMSO as described previously (19). Then the absorbance of the reaction mixture was measured spectrophotometrically at 415 nm. The total flavonoid content of the extract was expressed milligrams of quercetin equivalent (QE)/L of plant extract (mg QE/L), where the standard curve used was prepared with 0-100 mg/L of quercetin(Q) in DMSO.

Isolation of cytosol from bovine liver

The bovine liver were obtained from slaughterhouse of Kazan, Ankara, Turkey.

The liver samples were homogenized in 10mM potassium phosphate buffer (pH 7.0), containing 0.15M KCl, 1mM EDTA, and 1mM of DTT, by using a glass teflon homogenizer and centrifuged at 10,000 x g for 20 min. The supernatant was filtered through cheesecloth and the filtrate was centrifuged at 30,000 x g for 60 min. The collected supernatants were filtered again and the resultant filtrate was referred as cytosol (17). prepared homogenates, containing 46.41mg protein/mL, were kept in Hettich ultra low freezer (-80 °C) until used. The total protein content was determined by the Lowry method (20).

Inhibition of glutathione-S-transferase (GST) The change in total GST activities was measured against the substrate, 1-chloro-2,4dinitro -benzene (CDNB), by monitoring the thioether (GSH-CDNB conjugate) formation at 340 nm (21). The measurements were performed with cvtosol (0.982)protein/mL) and plant extracts (7 - 476 ng/mL) or control (DMSO alone), in 100 mM potassium phosphate buffer at pH 6.5 with 2.4 mM CDNB and 3.2 mM GSH, using optimized microplate application protocol (22).

Inhibition of glutathione peroxidase (GPx)

Glutathione peroxidase activity measured by previously reported method (23, 24) after some modifications for microplate applications (25). The GPx activity was measured against the substrate, tertiary butyl hydroperoxides (t-BuOOH), and the decrease nicotinamide adenine dinucleotide phosphate (NADPH) was monitored at 340 nm. The GPx activity changes were measured by using purified GPx $(37.5 \times 10^{-3} \text{ U/ml})$ and plant extracts (7 - 476 ng/mL) or control (DMSO alone), with 2 mM GSH, 0.25 mM NADPH, GSH-reductase (GR, 0.5 unit/ml) and 0.3 mM t-BuOOH, in 50 mMTrisHCl (pH 8.0). The reaction was initiated by adding GPx, and the change in absorbance was recorded at 340 nm for 5 min by using multimode microplate reader.

Inhibition of catalase (CAT)

Catalase (CAT) inhibition was determined by monitoring a red quinoneimine dye remaining hydrogen peroxide (26, 27). The assay was miniaturized for microplate application (28)and contained plant extraction solutions with final concentration was in the range of 2-285 ng/mL, 50 mM phosphate buffer (pH 7.0), 20 U/ mL purified bovine liver catalase, 0.0961 mM H₂O₂. The reaction was stopped by NaN3 and incubated at room temperature for 5 min, followed by the incubation with chromogen at room temperature for 40 min. Then, the absorbance was read at 520 nm, enzyme activity was calculated with respect to hydrogen peroxide remained which was determined calibration curve constructed in the range of 9.61 - 307.6 µM hydrogen peroxide.

Data analysis

The data analysis was performed using Graphpad Prism 6.0. The inhibitory activities of extracts against enzyme targets were calculated as 50 % inhibitory concentration, or IC₅₀ values, and obtained from doseresponse curves constructed. The enzyme calibration and the dose response curves were constructed using 2-3 independent experiments in 96 well microplates, each in duplicates or triplicates, using Multimode Microplate Reader (SpecraMax M2e, MDC, Sunnyvale, CA, USA).

RESULTS AND DISCUSSION

Each extract was prepared by dissolving 2g of dry samples in 20mL methanol. The methanol extraction of the flowers and leaves of *E. campestre* yielded 5.27% and 6.72% of dry weight, respectively.

The total phenolic contents were determined by using the Folin-Ciocalteu's method. According to the method, the total phenolic contents of extracts were calculated using the equation obtained from the standard curve of gallic acid graphic (y = 0.0749x + 0.0866, R^2 = 0.9898). The amount of total phenolic compounds found in the methanol extract from flowers of E. campestre was 116.69 mg GAE/L of plant extract. The total phenolic content in the leaves was 109.62 mg GAE/L of plant extract. The total concentration of flavonoids in extracts were determined by employing aluminum the chloride colorimetric method. The total flavonoid contents of extracts were calculated using the equation got from the standard curve of quercetin graphic (y = 0.0429x + 0.153, R² = 0.998). Total amount of the flavonoid contents found in the methanol extracts from flowers and leaves of E. campestre were 110.58 and 108.37 mg QE/L of plant extract, respectively. GSTs activity was determined against the substrate, 1-chloro-2,4-dinitrobenzene (CDNB), by monitoring the thioether formation at 340 nm. In order to calculate the percent inhibition of GST activity and IC50 values, the final concentration of plant extracts were used in the assay, was within the range of 7- 476 ng/mL. In this study, the inhibitory effect of the extracts obtained from *E.campsetre* flowers and leaves investigated and as it can be seen in the Figure 1, the flowers extract show the better inhibitory effect on GST enzyme with IC₅₀ value of 363 ng/mL than the leaves. In this study none of the extracts evaluated were shown any reasonable GPx and CAT inhibition.

In this study, we were concerned by the evaluation of phenolic and flavonoid contents of methanolic extracts of different parts of E. campestre to determine their inhibitory glutathione-Spotential the on transferase, glutathione peroxidase and enzymes activities. Phenolic catalase compound are having at least one or more aromatic rings with one or more hydroxyl attached (29).Many groups compounds have been reported to have potentials of antioxidant, anticancer, antiatherosclerotic, antibacterial, antiviral and anti-inflammatory activities (30).literature data show that plant extracts with high amount of phenolic compounds are known to have important inhibitory potential on glutathione-S-transferase (17,31,32). Thus, it can be play an important role in developing multi-drug resistance to chemotherapy in tumor cells. In present study, E. campestre flower extract has higher phenolic contents and exhibited expressive effect on the GST inhibitory activity.

Nebija et al. (33) studied mineral composition and *in vitro* antioxidant activity of the aqueous and ethanol extracts of aerial

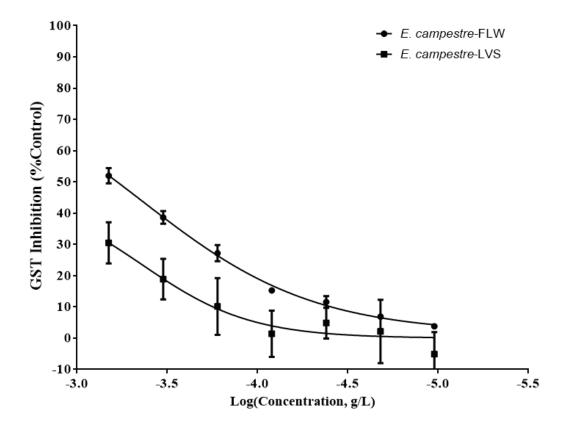


Figure 1. Percent GST inhibitory activity of methanol extract of *E. campestre* flowers (FLW) with IC₅₀ values of 363 ng/mL and methanol extract of *E. campestre* leaves (LVS).

and root parts of E. campestre by DPPH assay, inhibition of production of hydroxyl radical, β-carotene-bleaching assay, inhibition of lipid peroxidation (TBA test). They showed that, different parts of E. campestre accumulate different amounts of Zn, Fe, Cu, Mn, Ni, K, Co, Pb, Cd and Cr minerals. Also, the results obtained from evaluation of radical scavenging and antioxidant activity exhibited that higher radicalscavenging activity against DPPH- radical presented by the ethanol extract of root of E. campestre compared to the aerial part of the plant. Furthermore, the inhibition capacity on the production of hydroxyl radical in deoxyribose system was found to be strong (50% and 45% for aerial part and root ethanol extract, respectively). However, both ethanol extracts of E. campestre from aerial part and root, exhibited low antioxidant activity in βcarotene/linoleic acid system as well as low capacity for inhibition of lipid peroxidation in rat liver homogenate.

In another research, Kholkhal et al. (34) reported total phenolic and flavonoid content and antioxidant activity of methanol, acetone, ethyl acetate, and butanol extracts of *E. maritimum* roots. They determined that the amount of total phenolic and flavonoid contents were higher in acetone extract than methanol extract. Also, they observed that butanol extract exhibited higher free radical scavenging and iron reducing potential.

As a conclusion, this is the first study on the biological potential of E. campestre on the glutathione-S-transferase, glutathione peroxidase and catalase inhibitory activities. In this study we found that the flowers extract of E. campestre having higher phenolic content, were also more effective GST inhibitors with an IC₅₀ of 363 ng/mL.

ACKNOWLEDGEMENTS

This study was supported by the grant from The Coordination of Scientific Research Projects of Ankara University awarded to Prof. Dr. Özlem Yıldırım (Grant No: 13L4240008).

REFERENCES

- Duke JA, Bogenschutz-Godwin MJ, duCellier J, Duke PAK, Handbook of medicinal herbs, 2nd Ed, pp. 277-278, London, CRC Press, 2002.
- 2. Erdelmeier CAJ, Sticher O, A cyclohexenone and a cyclohexadienone glycoside from *Eryngium campestre*, Phytochemistry 25,741-743, 1986.
- Wörz A, On the distribution and relationships of the South-West Asian species of *Eryngium* L. (Apiaceae-Saniculoideae), Turk J Bot 28, 85-92, 2004.
- 4. Baytop T, Türkiye'de Bitkilerle Tedavi-Geçmisten Bugüne (Therapy with Medicinal Plants in Turkey-Past and Present) 2nd Ed, pp.169, Nobel Tip Basimevi, Istanbul, 1999.
- 5. Güner A, Özhatay N, Ekim T, Baser KHC, Flora of Turkey and the East Aegean Islands, Vol 11, pp.136-138, Edinburgh University Press, Edinburgh, 2000.
- Sezik E, Yesilada E, Tabata M, Honda G, TakaishiY, Fujita T, Tanaka T, Takeda Y, Traditional medicine in Turkey VIII. Folk medicine in East Anatolia; Erzurum, Erzincan, Agri, Kars, Igdir provinces, Eco Bot 51, 195-211, 1997.
- Kartal M, Mitaine-Offer AC, Asaker M, Miyamoto T, Calis I, Wagner H, Lacaille-Dubois MA, Two new triterpenesaponins from Eryngium campestre, Chem Pharm Bull 53(10), 1318-1320, 2005.
- Kartal M, Mitaine-Offer AC, Paululat T, Abu-Asaker M, Wagner H, Mirjolet JF, Guilbaud N, Lacaille-Dubois MA, Triterpenesaponins from *Eryngium campestre*, J Nat Prod 69(7),1105-1108, 2006.
- 9. Burg D, Mulder GJ, Glutathione conjugates and their synthetic derivatives as inhibitors of glutathione-dependent enzymes involved in cancer and drug resistance, Drug Metab Rev 34, 821-863, 2002.
- 10. Nabekura T, Overcoming multidrug resistance in human cancer cells by natural compounds, Toxins 2(6), 1207-1224, 2010.
- Sheehan D, Meade G, Foley VM, Dowd CA, Structure, function and evolution of glutathione transferases: implications for classification of non-mammalian members of an ancient enzyme superfamily, Biochem J 360, 1-16, 2001.
- 12. Potapova O, Haghighi A, Bost F, Liu CT, Birrer MJ, Gjerset R, Mercola D, The Jun kinase stress-activated protein kinase pathway

- functions to regulate DNA repair and inhibition of the pathway sensitizes tumor cells to cisplatin, J Biol Chem 272(22), 14041-14044,1997.
- 13. Tan AC, Konczak I, Sze DMY, Ramzan I,Molecular pathways for cancer chemoprevention by dietary phytochemicals, Nut Cancer Internat J 63(4), 495-505, 2011.
- 14. Townsend DM, Tew KD, The role of glutathione-S-transferase in anti-cancer drug resistance, Oncogene 22(47), 7369-7375, 2003.
- 15. Prasad KN, Cole WC, Kumar B, Prasad KC, Pros and cons of antioxidant use during radiation therapy, Cancer Treat Rev 28(2), 79-91, 2002.
- 16. Simone CB, Simone NL, Simone V, Simone CB, Antioxidants and other nutrients do not interfere with chemotherapy or radiation therapy and can increase kill and increase survival, Alter Therap Health Med 13(1), 22-28, 2007.
- 17. Coruh N, Sagdicoglu Celep AG, Ozgokce F, Iscan M, Antioxidant capacities of *Gundelia tourneforii* L. extracts and inhibition on glutathione-S-transferase activity, Food Chem 100, 1249-1253, 2007.
- Slinkard K, Singleton VL, Total phenol analyses: Automation and comparison with manual methods, J Enol Viticul 28, 49-55, 1977.
- 19. Chang CC, Yang MH, Wen HM, Chern JC, Estimation of total flavonoid content in propolis by two complementary colorimetric methods, J Food Drug Anal 10(3), 178-182, 2002.
- 20. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ, Protein measurement with the Folin phenol reagent, J Biol Chem 193(1), 265-275, 1951.
- 21. Habig WH, Pabst MJ, Jakoby WB, Glutathione-S-transferases the first enzymatic step in mercapturic acid formation, J Biol Chem 249, 7130-7139, 1974.
- 22. Isgor BS, Isgor YG, Effect of alpha-1-adrenoceptor blocker on cytosolic enzyme targets for potential use in cancer chemotherapy, Int J Pharmacol 8(5), 333-343, 2012.
- 23. Paglia DE, Valentine WN, Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase, J Lab Clin Med 70(1), 158-169, 1967.
- 24. Yıldırım Ö, Büyükbingöl Z, Effects of supplementation with a combination of cobalt and ascorbic acid on antioxidant enzymes and lipid peroxidation levels in streptozotocindiabetic rat liver, Biological Trace Element Research 90 (1-3), 143-154, 2002.

- 25. Isgor BS, Isgor YG,Ozalp-Yaman S, The role of metal coordination complexes in cytosolic cellular defense, Pure Appl Chem 85(2), 365-375, 2013.
- 26. Bai JZ,Saafi EL, Zhang SP, Cooper GJS, Role of Ca²⁺ in apoptosis evoked by human amylin in pancreatic islet beta-cells, Biochem J 343, 53-61, 1999.
- 27. Yıldırım Ö, Aras S, Ergül A, Response of antioxidant system to short-term NaCl stress in rootstock and razaki cultivar, Acta Biol Cracoviensia Series Bot 46, 151-159, 2004.
- 28. IsgorYG, Iscan M, Ozturk HS, Durak I, Effects of water soluble garlic extract on human leukemia HL60 cell lines, Turk J Biochem 33(3), 78-84, 2008.
- 29. Fresco P, Borges F, Diniz C, Marques MP, New insights on the anticancer properties of dietary polyphenols, Med Res Rev 26(6), 747-766, 2006.
- 30. Huang WY, Cai YZ, Zhang Y, Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention, Nutr Cancer 62(1), 1-20, 2010.
- 31. Zhang K, Wong KP, Glutathione conjugation of chlorambucil: measurement and modulation

- by plant polyphenols, Biochem J 325, 417-422, 1997.
- 32. Gyamfi AM, Ohtani II, Shinno E, Aniya Y, Inhibition of glutathione-S-transferases by thonningianin A, isolated from the African medicinal herb, *Thonningia sanguinea* in vitro, Food Chem Toxicol 42, 1401-1408, 2004.
- 33. Nebija F, Stefkov G, Karapandzova M, Stafilov T, KadifkovaPanovska T, Kulevanova S, Chemical characterization and antioxidant activity of *Eryngium campestre* L., Apiaceae from Kosovo, Macedon Pharm Bull 55 (1, 2), 22-32, 2009.
- 34. Kholkhal W, Ilias F, Bekhechi C, AtikBekkara F, *Eryngium maritimum*: A rich medicinal plant of polyphenols and flavonoids compounds with antioxidant, antibacterial and antifungal activities, Curr Res J Bio Sci 4(4), 437-443, 2012.

Received: 06.03.2014 Accepted: 08.05.2014