

Proniosome: A Promising Approach for Vesicular Drug Delivery

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ABSTRACT

Different types of drug delivery systems are intended to deliver therapeutic agents to the appropriate site of interest to get desired pharmacological effect. In the field of drug delivery, the advancement of nanotechnology helps develop novel dosage forms such as liposome, noisome, and proniosome. Proniosomes are promising drug carriers, that are dry formulations, and after hydration, are converted to noisome dispersion. Dry proniosomal powder can deliver a unit dose of the drug with improved drug stability and solubility. By using this formulation, both the hydrophilic and lipophilic drugs can be delivered through different routes like oral, topical, transdermal, vaginal, *etc.* This review revolves on different features of proniosomes such as their structure, formulation materials of proniosomes, preparation methods, evaluation, and application. **Key words:** Proniosomes, vesicular drug delivery, evaluation of proniosome, application of proniosome

INTRODUCTION

A novel drug delivery system that delivers drugs at a preset rate set as *per* the need, pharmacologic aspects, drug profile, physiological conditions of the body, *etc.* In current times, no single drug delivery system meets all the standards; however, efforts are created through novel approaches.

The aim of novel drug delivery systems is targeted and controlled drug delivery. Colloidal delivery and nanotechnology have attracted the most interest because of promising systems for having a localized result.¹

The delivery of drugs using colloidal particulate carriers such as proniosomes is dry and free-flowing preparation coated with a surfactant. To form a multi-lamellar noisome, proniosomes are rehydrated directly within minutes by transient agitation. Niosome suspension is appropriate for giving medication by different routes. They are promising candidates for industrial applications as they can transport, distribute, store, and process easily. Therefore, proniosomes can be another alternative to liposomal and other vesicular drug delivery systems for the entrapment of both polar and non-polar medications.² Proniosomes improve effectivity, scale back or eliminate adverse effects and enhance therapeutic actions of medicine. They are accustomed to avoid the gastrointestinal tract incompatibility, pre-systemic metabolism, and unwanted adverse effects related to oral delivery. Additionally, they maintain therapeutic levels of drug for an extended time, decrease the frequency of administration and improve patient compliance.³⁻⁵

This article in brief reviews the types, fabrication, characterization, and pharmaceutical applications of proniosomes.

Structures of proniosomes

Proniosomes are microscopic lamellar structures, hexangular structures, and blackish structures, where their location is clear, semi-transparent, and semi-solid gel-like structures (Figure 1). Consistent with their methodology of preparation, proniosomes are unilamellar or multi-lamellar. They even have bilayer in their structure having hydrophilic ends that are exposed on the surface and hydrophobic chains that face one another within the bilayer inside the vesicles. Bilayer consists of non-ionic surface-active agents. To create a bilayer surfactant molecule,

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offers direction in such a way that hydrophilic ends of the nonionic surfactant are arranged toward the outside, whereas the hydrophobic ends exist in the opposite direction. Hydrophilic drugs are placed at intervals in the area encircled within the vesicle and the hydrophobic medication is implanted within the bilayer. For association, in liquid media, proniosomes attach to cholesterol with different categories of non-ionic surfactant like alkyl radical or dialkyl polyglycerol ether.⁶

Materials used for the preparation of proniosomes

Surfactant: Surfactants, especially non-ionic surfactants are the key structural components in the preparation of proniosomes. These surfactants do not have any charge as they possess a polar head and non-polar tail. So, their stability, toxicity and compatibility is higher than other surfactants. The non-ionic surfactants have wet and emulsifying effects by which they improve the solubility and permeability of drugs. The hydrophilic-lipophilic balance (HLB) value is critical for selecting surfactants and HLB value between 4 and 8 is compatible with vesicle formation by proniosomes. It is difficult for hydrophilic surfactants to achieve a high concentration because of the high liquid solubility of hydrophilic surfactants. Therefore, aggregation and conglutination to form a proniosomal lamellar structure would be absent (Table 1).⁷

Cholesterol: Cholesterol can interact with non-ionic surfactants and regulates the physical and structural properties of

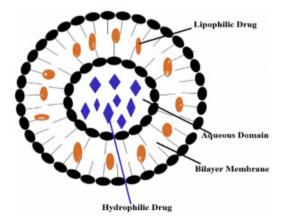


Figure 1. Structure of a proniosome

Table 1. Commonly used surfactants with their HLB values

proniosomes.⁸ It improves the stability and rigidity of the proniosomal membrane and controls drug permeation through the membrane. Depending on the HLB value of the surfactants, the amount of cholesterol required for the preparation of proniosomes is determined. When the HLB value is above 10, the amount of cholesterol to be increased to cover the larger groups.⁹ But entrapment efficiency (EE) of the prepared formulation is decreased¹⁰ above a certain level of cholesterol, possibly due to a decrease in volume diameter.¹¹

Lecithin: Lecithin is a phospholipid that acts as a membrane stabilizer in the formulation of proniosomes. The most common lecithins that are used in the formulation are soya and egg lecithin and it has been reported that hydrogenated-type lecithins have advantages over not hydrogenated lecithins, give increased rigidity of the cholesterol and help in the formation of tight vesicles.¹² Double bonds in non-hydrogenated lecithin allow the molecular chains to bend (conformational rotation), which prevents tight contact with the adjacent molecules on forming the niosomal membrane. This results in low rigidity and high permeability of the membrane.

Hydration medium: Generally, the hydration medium used in proniosomes is phosphate buffer. Depending on the solubility of the encapsulated drug, the pH of the buffer is selected.¹³ Ruckmani and Sankar¹⁴ ascertained that drug leakage increased with the increase in the volume of hydration medium but simultaneously, EE increases, when the hydration time was increased from 20 to 45 min.

Organic solvent: The solvent can act as a penetration enhancer. It also greatly affects the size of the vesicles formed. The size of the vesicle and permeation rate of the drug in a proniosomal formulation are influenced by the type of alcohol. Different sized vesicles are formed using different alcohols as they have the order: » isopropanol < butanol < propanol < ethanol.¹⁵

Carrier material: Carrier materials accommodate the drug in the proniosomal formulations. Carriers should have safe, non-toxicity, free-flowing properties. They should possess low solubility in the solution of loaded, but good solubility in water for ease of hydration. They increase the surface area and impart flexibility to the proniosomes. The frequently used carrier materials are sorbitol, mannitol, maltodextrin, glucose monohydrate, spray-dried lactose, sucrose stearate, and lactose monohydrate.¹⁶

Name of surfactant	M.W. (g/mol)	HLB value	Transition temperature
Span 20 (sorbitan monolaurate)	346.46	8.6	16°C
Span 40 (sorbitan monopalmitate)	402.57	6.7	42°C
Span 60 (sorbitan monostearate)	430.63	4.7	53°C
Tween 20 (polyoxyethylene sorbitan monolaurate)	522.68	16.7	-
Tween 60 (polyoxyethylene sorbitan monostearate)	648.92	14.9	-
Tween 80 (polyoxyethylene sorbitan monooleate)	604.82	15.0	-
Span 85 (sorbitan trioleate)	957.49	1.8	-

M.W.: Molecular weight, HLB: Hydrophilic-lipophilic balance

Preparation method of proniosomes

A drug that has poor aqueous solubility, low bioavailability and dissolution, poor membrane permeability, low absorption profile, excessive metabolism, variable plasma concentration, and poor patient efficiency is suitable to encapsulate into proniosomes.¹⁷ Three methods are available for proniosomal drug formulation (Figure 2).

Slurry method

In this method, a single or a mixture of organic solvent is used in the preparation of a stock solution of surfactant and membrane stabilizer. The drug and carrier are dissolved in a membrane stabilizer solution and all the components are mixed until a slurry is formed. With the help of a rotary evaporator at specified conditions (*e.g.* 50-60 rpm, $45 \pm 2^{\circ}$ C temperature, and 600 mm of Hg pressure), the slurry is dried, and the freeflowing product is obtained. The obtained free-flowing dried material is further dried with the help of a desiccator at room temperature under vacuum to get proniosomes.¹⁸

Slow spray coating method

The slow spray-coating method is carried out by spraying organic solution, surfactant, cholesterol, and drug onto the carrier and then removing the solvent using a rotary evaporator under controlled conditions at 65-70°C for 15-20 min. Until the desired surfactant loading has been achieved, the process is continued and repeated. The vaporization should be carried on until the powder becomes completely dry.^{19,20}

Coacervation phase separation method

Most of the proniosomal gel (PNG) is prepared by this method. In this method, exactly measured amounts of drugs, surfactants, and cholesterol are placed in a clean and dry glass vial having a wide opening. Then, the solvent is added and warmed in a water bath at 60-70°C until the surfactant and cholesterol is fully dissolved. To prevent the evaporation of the solvent, the open end of the vial should be covered with a lid. Followed by the addition of an aqueous phase in the vial, the mixture was warmed in the water bath to get a clear solution. It is then cooled at room temperature, between this time PNG is produced from the dispersion.²¹

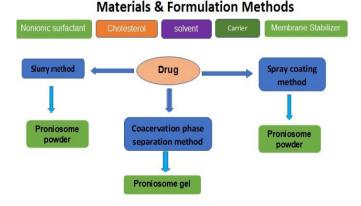


Figure 2. Schematic representation of materials and formulation methods of proniosomes

Evaluation of proniosomes

A group of properties of proniosomes can be evaluated using different methods. These methods are described below.

Vesicle size and shape

On hydration, proniosomes are converted to globular-shaped niosomes. The size and morphology of the niosomes can be determined by optical microscopy, scanning electron microscopy (SEM), photon correlation microscopy, transmission electron microscopy (TEM), and freeze-fracture electron microscopy.²² Dynamic light scattering (DLS) method is applied to measure the vesicular size distribution. DLS essentially measure fluctuations in scattered light intensity due to diffusing particles, the diffusion coefficient of the particles can be determined. The vesicle size of the noisome is measured without agitation and with agitation. If hydration is performed without agitation, biggest size is formed.²³

Angle of repose measurement

The angle of repose of dried proniosomes prepared using the slurry and spray coating method is measured by the funnel and cylinder technique.

Zeta potential (ZP)

The stability of the particle can be ensured with the value of ZP. This is ascribed to the electrostatic repulsion between particles with the same electric charge that causes the segregation of the particles. A high ZP value leads to increased repulsive interactions in charged particles and prevents the agglomerate formation between the particles. This ensured uniform size distribution in proniosomes. A proniosomal formulation having ZP value minimum \pm 30 mV is considered a physically stable formulation. So, aggregation of particles can be avoided.^{24,25}

Osmotic shock

An osmotic shock study helps in the determination of vesicle size changes. For this, the proniosomal formulations are incubated in different types of solution like hypertonic, isotonic, and hypotonic solutions for 3 h. Changes in vesicle size are detected by an optical microscope.²³

Entrapment efficiency (EE)

To study EE, we should separate the free drug using several techniques like dialysis, gel filtration, ultracentrifugation, column chromatography, and freeze-thawing. Two techniques can be applied to measure EE. One is the destruction of the proniosomal vesicle with propane (50%) or triton (0.1%) and the entrapped drug is determined. Another method is that after the destruction of the vesicle, the un-entrapped drug is measured.²⁶

The percentage of entrapment was calculated using the following formula:

% Entrapment=
$$\frac{ED}{TD}$$
 X 100

where ED is the amount of entrapped drug and TD is the initial amount of drug.

The calibration curve is used to calculate drug content. For this, proniosomes are lysed with methanol in a volumetric flask by shaking for 15 min. Then, the stock solution is prepared with methanol. With the help of phosphate buffer, 10% solution is prepared from the stock solution. Aliquots are withdrawn and absorbance is measured followed by a drawing of calibration curve.²⁴

Particle size (PS) measurement

One of the important criteria for prepared proniosomes is PS. With the help of SEM, the uniformity of PS and surface characteristics is measured. Optical microscopy is used to confirm the formed vesicles after hydration of proniosome.²⁶

Rate of spontaneity

The rate of spontaneity is the measure of the number of niosomes formed following hydration of proniosomes. To determine the rate of spontaneity, PNG is transferred and spread uniformly along the walls of the small stoppered glass tube container. Then, NaCl (0.154 M) was added with caution and placed to one side without any turbulence. With the help of Neubauer's chamber, the number of niosomes eluted from proniosomes is calculated.²⁷

In vitro dynamics study

The *in vitro* release study is a critical test to assess the safety, efficacy, and quality of nanoparticle-based drug delivery systems. *In vitro*, drug release kinetics of the prepared proniosomes can be determined using a Franz diffusion cell, Keshary-Chien diffusion cell, dialysis membrane, reverse dialysis, and United States Pharmacopeia dissolution apparatus type 1.²⁸

Stability study

To ensure the stability of the prepared proniosomes, they are placed at a various temperatures, freezing temperature (2-8°C), normal temperature (25 ± 0.5°C) and elevated temperature (45 ± 0.5°C) for 1-3 months and the change in drug content and mean vesicle diameter is observed at a different time interval. The International Conference on harmonization (ICH) guidelines propose dry proniosomes powder should be studied for the accelerated stability at 75% relative humidity and 40°C as *per* international geographical zones and geographical conditions.²⁹

Proniosomal drug delivery through different routes

Oral routes

The oral route of drug administration is the most preferred route for drug delivery. But bioavailability of the orally administered drug is sometimes affected by first-pass metabolism, instability in the gastric environment, low permeability through the intestinal epithelium. In some cases, absorption of the drug may be altered due to the presence of food. Thus, to improve the bioavailability of the oral drug, different nanocarriers are engaged. Oral proniosomes are one them that can solve the limitations of the conventional oral dosage form.³⁰ *In vitro* release kinetics of oral vinpocetine (VP) prepared using the slurry method indicated a

faster release rate of reconstituted niosomes in contrast to VP suspension at pH 6.8 or 7.2 phosphate-buffered saline. In vivo pharmacokinetic study data also showed a better correlation with the *in vitro* data.³¹ Oral acemetacin also prepared using the slurry method in proniosomal powder and tablet formulations, displayed better pharmacokinetic properties.³² Lornoxicam is a widely used analgesic drug that belongs to the non-steroidal anti-inflammatory group. Proniosomal form of lornoxicam showed significantly higher (p<0.05) transmucosal flux across the oral mucosa than lornoxicam containing carbopol gel and the diffusion of lornoxicam was higher (more than two folds) in proniosomal formulation.³³ Proniosomal telmisartan tablets prepared with surfactants having different HLB values (span 40 and brij 35), cholesterol (20-50%), and phospholipids (egg yolk and soybean). In vitro as well in vivo comparative study showed extended drug release with a higher $\rm C_{max}$ The $\rm C_{max}$ was increased 1.5 fold while AUC and also increased significantly 3 fold compared with the commercial tablet. The sustained release pattern of telmisartan was indicated by t_{max}, which was increased 3 fold in contrast to conventional tablets. The relative bioavailability was also increased by 3.2 fold (Table 2).34

Ocular routes

Proniosomes are one of the promising methods in ocular drug delivery. In this route, PNGs in the ocular route provide several advantages like extended and sustained action, enhanced corneal residence time, prevented enzymatic degradation of drugs in tears, and ultimately improve ocular bioavailability.35 Lomefloxacin HCl prepared using the coacervation phase separation method. The results showed that the area under the curve (AUC) of proniosomal formulation is higher (722.45 ± 0.01) than the marketed product (126.25 ± 0.049) .³⁶Li et al.³⁷ developed proniosomal formulation of tacrolimus, an immunosuppressive agent for topical ophthalmic delivery containing lecithin and poloxamer 188 as surfactants, cholesterol. In vitro studies in rabbit cornea of stable tacrolimus-loaded proniosomes showed enhanced precorneal permeation and retention of tacrolimus. The in vivo ocular irritation test of 0.1% tacrolimus-loaded proniosomes in rat eyes for 21 successive days (four times in a day) showed no irritation and good compatibility with the cornea. Proniosomes were also found to prolong the survival of corneal grafts and showed practical corneal anti-allograft rejection efficacy in the xenotransplantation model. Curcumin-loaded PNG was developed for treating ocular inflammation with the help of cremophore RH, lecithin, and cholesterol. Selected PNG showed 3.22 fold and 1.76 fold higher permeability rates than curcumin dispersion and its freeze-dried form, respectively.38 This formulation could be an effective, better biofriendly alternative for the therapy of inflaming eyes.

Emad Eldeeb et al.³⁹ studied on brimonidine tartrate (BRT) proniosomal formulation used in glaucoma developed using the coacervation phase separation method. They used two surfactants having higher and lower HLB values, namely tween 80 (HLB 15) and span 60 (HLB 4.7), respectively. The results of BRT PNG showed a 7.90 fold increase in mean residence time compared to the marketed product.

Route	Preparation method	In vitro/in vivo effects	Reference
Oral vinpocetine (VP)	Slurry method	VP-loaded proniosomes showed a significant improvement in the bioavailability	31
Oral acemetacin	Slurry method	Both proniosomal powder and tablet formulations showed better pharmacokinetic properties	32
Oral lornoxicam	Proliposome-liposome method	The transmucosal flux of proniosomal formulations lornoxicam across the oral mucosa was significantly higher (p <0.05) than lornoxicam containing carbopol gel and the percent drug diffused increased more than two folds	33
Oral telmisartan tablets	Slurry method In vitro as well in vivo comparative study showed extended drug release		34
Ocular lomefloxacin HCl	Coacervation phase separation method	The results showed that the area under the curve of proniosomal formulation is higher than the marketed product	36
Ocular tacrolimus	The proliposome-liposome method	Showed enhanced precorneal permeation and retention of tacrolimus	37
Curcumin-loaded ocular proniosomal gels	Coacervation-phase method	Proniosomal gel showed enhanced permeability	38
Ocular Brimonidine tartrate (BRT)	Coacervation phase separation method	The <i>in vivo</i> pharmacodynamic study confirmed the improved ocular bioavailability of BRT selected formula compared with marketed products	39
Ocular dorzolamide-HCl	Coacervation phase separation method	<i>In vivo</i> performance of optimized dorzolamide-HCl showed sustained effect and a significant reduction in intraocular pressure	40
Cromolyn sodium nebulizer	Proniosomes were prepared according to the method developed by Hu and Rhodes ²	High nebulization efficiency percentage and good physical stability	26
Aerosol beclometasone dipropionate (BDP) niosomes	Thin-film method	Entrapment of BDP in proniosome-derived niosomes was higher than that in conventional thin film-made niosomes	
Vaginal tenofovir disoproxil fumarate	Coacervation phase separation	The results showed a controlled and sustained release rate	44
Vaginal terconazole	-	Results indicated that the selected formula showed good stability and provided higher mucoadhesion and retention time then the commercial product, which resulted in more efficient <i>in vitro</i> inhibition of <i>Candida</i> <i>albicans</i>	45
Parenteral flurbiprofen	Slurry method	Sustained anti-inflammatory activity and decreased dosing frequency	46
Parenteral letrozole (LTZ)	Slurry method	Drug release exhibited a biphasic pattern, being fast at the first 24 h (up to 65% released) followed by a very slow-release phase for one month, releasing at least 95%. Overall, in this study, a facile approach to generating niosomes incorporating LTZ using a slurry- based proniosome technology was demonstrated	47
Transdermal piroxicam	Coacervation phase separation method	Maximum flux achieved was 35.61 µg/cm²/h, an enhancement of 7.39 times achieved for the transdermal system based on proniosomal gel as compared to control gel	48

Route	Preparation method	In vitro/in vivo effects	References
Transdermal ketoprofen	Slurry method	Showed significantly higher cumulative amount of drug permeated and steady-state transdermal flux compared to plain gel	49
Transdermal meloxicam	Coacervation phase separation method	The prepared proniosomes significantly improved drug permeation and reduced the lag time (p (0.05). Proniosomes prepared with span 60 provided a higher meloxicam flux across the rat skin than those prepared with tween 80. Testing of the anti-inflammatory effect of meloxicam proniosomal gel showed better pharmacological activity compared with the standard meloxicam gel	50
Transdermal celecoxib	Coacervation phase separation method	The selected proniosomal gel produced 100% inhibition of paw edema in rats up to 8 h after carrageenan injection. It produced 95% and 92% inhibition after 12 h and 24 h, respectively	51
Transdermal tenoxicam	Coacervation phase separation method	The investigated tenoxicam loaded proniosomal formula proved to be non-irritant, with significantly higher anti- inflammatory and analgesic effects compared to that of the oral market tenoxicam tablets	30
Transdermal fluconazole	Coacervation phase separation method	The results showed a well-defined spherical vesicle with sharp boundaries. Moreover, this formulation showed an excellent microbiological activity represented by a greater zone of inhibition (5.3 cm)	52
Transdermal estradiol	Coacervation phase separation method	The proniosome gel provided a higher permeation flux of estradiol across the skin compared with the noisome suspension	53
Dermal boswellic acid	Coacervation phase separation method	<i>In vitro</i> , skin permeation study showed the most sustained release in 24 h	54
Transdermal atorvastatin Coacervation phase separation calcium (ATC) method		Results suggest a promising, easy-to-manufacture, and effective ATC proniosomal gel for the safe treatment of hyperlipidemia	55
Transdermal cilostazole (CLZ)	Coacervation phase separation method	The CLZ-loaded proniosomes showed promising results with high potential to delivery it across the skin	56
Transdermal CLZ	Coacervation phase separation method	The results suggested that clozapine could be effectively loaded into proniosomal gel for administration through the skin	57
Transdermal galantamine hydrobromide	Coacervation phase separation method	The <i>in vitro</i> drug diffusion studies revealed that the proniosomal gel containing tween 80 showed maximum drug diffusion (99.24%) and the gel containing span 20 showed minimum drug diffusion (71.74%)	58
Intranasal duloxetine	Coacervation phase separation method	Result showed significantly improved permeation enhancement and stability with better control over drug release for a longer period through intranasal administration	54

Simultaneously, the relative area under the plasma concentration-time curve over the last 24 h dosing interval (AUC₀₋₂₄) value was also increased to 5.024 folds. Draize test also ensured that the formulations had no ocular irritation.³⁹ Dorzolamide-HCl (Dorz) is an anti-glaucoma drug soluble in water. The PNG of this drug was developed to sustain its effect

and to reduce dosing frequency. Here, the gel was developed by using L- α -lecithin, cholesterol, span 40 using the coacervation phase separation method. *In vivo* results showed a maximum reduction in intraocular pressure (IOP) of 32.6 ± 2.7 at 1.5 h. On the contrary, the PNG showed a sustained decrease in IOP with a maximum value of 45.4 ± 8.2 at 6 h, which was significantly

higher than standard formulation. Even after completion of the experimental period (8 h), the % dec in IOP was 19.5 \pm 9.2. This implies a prolonged release of Dorz PNG from the optimized formulation.⁴⁰

Pulmonary routes

With the aid of the pulmonary route, one can easily treat respiratory diseases than other delivery methods. Through this route, drugs can be directly applied within the lungs. Drug-loaded particles like liposomes dispensed through aerosol can easily distribute to the bronchi and lungs and prolong the release of the drug. Liposomal delivery also has minimum systemic side effects due to localized action to the lungs. But liposomes may be degraded by oxidation or hydrolysis. So, the proniosome can be an option to overcome the limitations of the liposome.⁴¹

For pulmonary drug delivery, the air-jet nebulizer is known very well. Proniosome-derived niosomes of cromolyn sodium were prepared by Abd-Elbary et al.²⁶ They used sucrose stearates in the formulation. The results exhibited a controlled release of drugs from the proniosome-derived niosomes compared to standard drug solution. Furthermore, high nebulization efficiency and physical stability were also achieved.²⁶

Likewise, aerosol properties of beclometasone dipropionate (BDP) niosomes using Aeroneb Pro and Omron Micro Air vibrating mesh nebulizers and Pari LC Sprint air-jet nebulizer were investigated by Elhissi et al.⁴² The study demonstrated that the satisfactory EE of BDP in proniosome-derived niosomes and the value was higher than that in conventional thin film-made niosomes.⁴²

Vaginal routes

Vaginal drug delivery is one of the favorable routes to target the disease associated with female health issues. It offers both the local and systemic delivery of drugs. Usually, different categories of drugs like antibiotics, antifungal, antiprotozoal, antiviral, labor-inducing agents, spermicidal agents, steroids, etc. are delivered through the vaginal route.43 PNG has excellent mucoadhesive properties and provides a constant release pattern, which is very useful for vaginal drug delivery. Tenofovir disoproxil fumarate (TDF) is an antiretroviral drug (a nucleotide analog) that works through the inhibition of viral reverse transcriptase. PNG of TDF was prepared with the help of cholesterol, surfactants (span 20, 40, 60, 80, tween 20 and 80), lecithin by coacervation phase separation method. A comparative in vivo dissolution study was conducted between proniosomes suppository, drug suppository and PNG formulations for 24 h using cellophane membrane, our results indicated the proniosomal suppository. Another result revealed a controlled and sustained release rate compared to the other two formulations.⁴⁴ Terconazole, an antifungal drug, PNGs were developed on the basis of span 60 and brij 76 in different molar ratios (1:1, 1:1.5, and 1:2) relative to cholesterol. The results displayed that increased concentration of cholesterol relative to the surfactant affected both EE and vesicle size of niosomes prepared by incorporating into 1% carbopol gel. Drug release

profiles from different prepared PNG formulations in simulated vaginal fluid studied in comparison with the commercial product of terconazole for 24 h. Depending on the high EE % and *in vitro* release profile, selected formulation was further evaluated for stability, mucoadhesion to the vaginal mucosa and inhibition of candida growth. Results indicated that the selected formula was in good stability and provided higher mucoadhesion and retention time than the commercial product, which resulted in more efficient *in vitro* inhibition of *Candida albicans*.⁴⁵

Parenteral routes

In parenteral drug delivery, targeted and sustained drug release at a predetermined rate can be achieved due to remarkable advancement in pharmaceutical technology. Flurbiprofen⁴⁶ and letrozole⁴⁷ are prepared by the slurry method. Both drugs showed sustained activity and reduced dosing frequency.

Dermal and transdermal routes

The dermal route is employed for local action only to treat different types of skin disease. This route can avoid systemic effects and therefore offers fewer side effects. However, through transdermal delivery, we can deliver drugs for systemic action. But in both the dermal and transdermal drug delivery, the skin prevents the penetration of drugs. Vesicular drug delivery can be used to overcome this problem.

Non-steroidal anti-inflammatory drugs (NSAIDs) such as piroxicam,⁴⁸ ketoprofen,⁴⁹ meloxicam,⁵⁰ celecoxib,⁵¹ and tenoxicam³⁰ are planned to avoid gastrointestinal adverse effects. Here, all the NSAIDs except ketoprofen are prepared by the coacervation phase separation method, whereas ketoprofen is prepared by the slurry method.

Fluconazole-loaded PNGs were prepared by the coacervation phase separation method using different non-ionic surfactants (spans and tweens). The prepared fluconazole PNGs were evaluated for various parameters such as PS, drug EE %, and in vitro drug release. The experimental results showed that the EE % for the prepared formulae are acceptable (85.14-97.66%) and they are size (19.8-50.1 nm). The planned gel also showed sustained drug release. The formulation, which was prepared from span 60:tween 80 (1:1), and cholesterol showed highest EE % and gave slow release (40.50 ± 1.50% after 6 h), was subjected to ZP test, TEM as well as microbiological study. The results indicated a well-defined spherical vesicle with sharp boundaries and good physical stability of fluconazole within the prepared gel. Moreover, this formulation showed an excellent microbiological activity represented by a greater zone of inhibition (5.3 cm) compared with control gel (fluconazole in 2% hydroxy propyl methyl cellulose gel formula) (4.2 cm) and plain gel with no drug (0 cm) against *C. albicans.*⁵² Fang et al.53 studied transdermal estradiol gel and the results provided a higher permeation flux of estradiol across the skin. In vitro skin permeation study of dermal boswellic acid gel, was studied for 24 h, and a sustained release pattern was observed (84.83±0.153 mg/cm²). Inhibition of inflammation of the proniosomal patch was also significantly (p<0.001) higher compared to the marketed gel at the same dose.⁵⁴

In an attempt to modify the anti-hyperlipidemic effect and to reduce statins-induced hepatotoxicity, atorvastatin calcium (ATC) transdermal PNG was developed by a coacervation phase separation method. Different non-ionic surfactants (spans, tweens, cremophor RH 40, and brij 52) were incorporated in the vesicle's lipid bilayer, along with lecithin. PNG gel was characterized for encapsulation EE %, vesicle size, polydispersity index (PDI), and ZP. The results revealed nano-sized (<350 nm) range vesicles with relatively high ATC EE (70.12-88%). Ex vivo results of the selected formulation demonstrated the permeation superiority of ATC proniosomes over free drugs. The selected PNGs showed significantly high flux ranging from 4.23 to 8.46 µg/cm² h - 1 with permeability coefficient values (P) (0.004-0.008 cm/h) when compared to free ATC dispersion, which significantly possessed lower flux and permeability coefficient results (2.92 µg/cm²h - 1 and 0.003 cm/h respectively). The pharmacodynamic study revealed that transdermal administration of ATC-PNG succeeded in retaining the antihyperlipidemic efficacy of orally administered ATC without elevating liver biomarkers. Histological examination signified the role of optimized ATC-PNG in hindering statininduced hepatocellular damage.⁵⁵ Transdermal cilostazole (CLZ) proniosomes were prepared by a coacervation phase separation technique. The optimum formula composed of 540 mg span 60 and 59.7 mg of cholesterol, had the highest EE % of (75.125 ± 0.125%), PS of (300.3 ± 0.2 nm), ZP of (-39.35 ± 0.15 mV), the percentage of the drug released after 2 h was (24.32 ± 0.13%) and after 24 h was (81.175 ± 0.325%). The safety of the proniosomes for topical application was confirmed by the histopathological examination. The CLZ-loaded proniosomes showed promising results with high potential to delivery it across the skin.⁵⁶ CLZ loaded PNG was prepared by the coacervation phase separation method using span 60, cholesterol, and lecithin. The optimized formulation had the highest EE of 90% and an average PS of approximately 325 nm PDI reflected homogeneity in the formulation. ZP was -59.76 mV, high enough to indicate a stable formulation. The in vitro release studies manifested a sustained release behavior of clozapine from the PNG. The ex vivo permeation demonstrated noteworthy permeation of the drug through stratum corneum with a steady state flux of 18.26 ug/cm²/hr.⁵⁷ Galantamine hydrobromide (HBr) is used for treating Alzheimer's disease and is described as proniosome gel by coacervation phase separation method to overcome the side effects of oral delivery. Microscopical observations of the gels showed vesicles of optimum size from 3.030 - 3.735 mm. The gel also showed an optimum rate of spontaneity in the range 9.60 mm³ x 1000 to 11.80 mm³ x 1000 and EE of vesicles in the range 66.15% to 86.92%. The gels had pH in suitable range of skin (5.92-6.9). The in vitro drug diffusion studies revealed that the PNG containing tween 80 showed maximum drug diffusion (99.24%), whereas the gel containing span 20 showed minimum drug diffusion (71.74%).58

Intranasal routes

The nasal drug delivery method has some limitations like mucociliary clearance, degradation of drugs by the enzyme.

Vesicular drug delivery systems can circumvent these limitations. Duloxetine (DX) is a new norepinephrine reuptake inhibitor used for treating depression. But it has high firstpass metabolism and low bioavailability (<50%) following oral administration, eventually leading to low cerebrospinal fluid concentrations. Khatoon et al.⁵⁹ designed mucoadhesive thiolated chitosan (TCS) gel containing proniosomes of DX for intranasal drug delivery to enhance the drug's contact time with nasal mucosa, bypass the first-pass effect and target the brain possibly using the olfactory pathway. Here, soya lecithin, cholesterol, and tween 80 was used in the preparation of the gel. pH of the DX-loaded proniosomal gel (D-MPNG) was 5.67 ± 0.145, indicating the compatibility of formulations within the nasal cavity without producing irritation. Notably, D-MPNG exhibited better control, releasing only 24% DX at pH 7.4 over 24 h compared to 78% release at pH 5.5. The presence of thiol groups of TCS significantly controls water uptake, resulting in moderate swelling and higher viscosity; thus, providing a sustained effect for a longer period.59

Cosmeceuticals application of proniosomes

Cosmeceuticals are generally used to refer to skincare products that contain active ingredients that are beneficial for improving the skin's appearance and promoting healthy skin.⁶⁰ Antiaging cosmeceuticals are most frequently recommended by physicians, who use them as an integral part of a comprehensive skin rejuvenation program. Moisturizers and serum containing ingredients such as vitamin C, niacinamide, retinol, peptides, growth factors, and botanicals can all be used in this regard. Additionally, patients undergoing cosmetic procedures such as laser resurfacing and chemical peels may be given cosmeceuticals to prime the skin for procedures, encourage healing, and reduce complications after. Cosmeceuticals are also recommended for patients with acne, rosacea, eczema, and other skin conditions, where they are commonly used along with prescription medications. For example, moisturizers containing anti-inflammatory botanical ingredients may be used in conjunction with prescription medications for treating rosacea. Cosmeceuticals containing soy can be used to provide added skin lightening benefits when paired with hydroquinone.

Applying therapeutic and cosmetic agents onto or through skin requires a non-toxic, dermatologically acceptable carrier, which not only controls the release of the agent for prolonging action but also enhances the penetration to the skin layer.⁶¹ Proniosome gel meets such criteria, which are useful for the delivery of cosmetics and cosmeceuticals. The therapeutic agents which can be used for incorporation into proniosomal carrier systems include, moisturizing, nutritional, anti-wrinkle, anti-aging, cleansing, sunscreen particles, *etc*.

Proniosome is a potentially scalable method to produce niosomes for the delivery of hydrophobic or amphiphilic drugs.⁶² Anti-aging cream containing the methanolic purple glutinous rice extract loaded in niosomes was developed by Manosroi et al.⁶³ Anthocyanin present in purple glutinous rice extract is responsible for the anti-aging activity. After 6 cycles of heating and cooling test, the formulation with 1% w/v of the purple glutinous rice extract contained 52.28% anthocyanin of

the initial. For *in vivo* antiaging activities, a cream containing niosomes loaded with the extract gave significantly decreased melanin index and skin roughness reduction of 14.05 and 9.95% of the initial, respectively. The percentage changes of the increased skin hydration, skin elastic extension, and skin elastic recovery when applied on human volunteers' skin with this formulation were +48.73, 24.51, and +35.98%, respectively

Tretinoin (TRT) is a widely used retinoid for the topical treatment of acne, photo-aged skin, psoriasis, and skin cancer. TRT-loaded proniosomes were prepared by the slurry method with the help of span 60 and D-sorbitol, span 40, cholesterol 95% stabilized, and tween 20.⁶⁴ prepared hydrated proniosomes were characterized by an evaluation of PS, the effect of drug concentration, EE, *etc.* EE of all hydrated proniosomal dispersions ranged from 76.6 ± 0.001% prepared to use span 40 to 94.15 ± 0.041% prepared using span 60. *In vitro* drug release was increased till 5th hour.

Diferuloylmethane or curcumin is obtained from turmeric, which possesses infiammatory properties blocking the formation of reactive oxygen species.⁶⁵ Proniosomes of curcumin were prepared using non-ionic surfactants (tween 80, span 60) either solely or in combination with cholesterol. The highest encapsulation efficiency of curcumin in niosomal formulations was 99.74%. Kinetically, niosomes fitted to the Korsmeyer-Peppas model with non-Fickian transport. The anti-inflammatory activity of curcumin in various formulations was evaluated using a rat hind paw edema method and the percentage of swelling was 17.5% following 24 h in the group treated with curcumin niosomal emulgel.⁶⁶

Coenzyme Q10 (CoQ10) also known as ubiguinone, an essential compound is found in every cell of the human body. CoQ10, an essential compound of cellular bioenergetics, also acts as a strong antioxidant and protects the body against aging.⁶⁷ CoQ10 proniosomes were prepared using the standard method with the help of Q10, span 85, soya lecithin, and cholesterol. In vitro drug release of CoQ10 followed a special cubic model, as the statistics of the chosen model were found significant (p=0.0006). The change in the levels of soy lecithin had a great impact and showed a synergistic effect on the release characteristics. A skin permeation study showed that the cumulative amount of CoQ10 permeated in 12 h was found to be 515.85, 463.25, and 507.49 mg/cm², respectively for selected PNG formulations. Animal skin was treated with UV radiation followed by treatment of PNG CoQ10 and conventional CoQ10 present in a gel base. The effectiveness of the treatment was evaluated based on biochemical estimation and histopathological studies. By using CoQ10 PNG formulation, levels of superoxide dismutase, catalase, glutathione, and total proteins were restored by 81.3%, 72.1%, 74.8%, and 77.1%, respectively to that of the control group. Histopathological studies revealed better protection of skin treated with CoQ10 PNG compared to free CoQ10. The prepared PNG did not interact with the normal histology and, hence, tolerated by the animal skin compared to conventional gel. Assessments of the formulations for various enzymatic and non-enzymatic estimations in animal skin after UV irradiation proved the efficacy of the developed formulation.

Proniosomal formulations containing the natural antioxidant resveratrol (*trans*-3,5,4'-trihydroxystilbene, RSV) were prepared by Schlich et al.⁶⁸ RSV is a polyphenol compound having anti-inflammatory,⁶⁹ neuroprotective,⁷⁰ anti-aging,⁷¹ and anticancer effects.⁷² Proniosomal powders were prepared by the slurry method and characterized. The hydration and sonication of proniosomes resulted in the formation of lipid nanoparticles with a mean diameter in the range 180-300 nm and a highly negative surface charge. RSV release from proniosome-derived niosomes was investigated in simulated gastric and intestinal fluid. biocompatibility assay carried out on intestinal cells (Caco2) demonstrated that proniosomes prepared with an HLB of 13.5 were significantly less toxic than their HLB16.7 counterpart. All the tested formulations could be employed safely at the doses commonly administered by the oral route.

Canthaxanthin (CTX) is a xanthophyll (a subclass of carotenoids) with widespread applications in pharmaceutical and cosmetic industries. It is a superior antioxidant and scavenger of free radicals compared with carotenoids such as β -carotene.⁷³ CTX was encapsulated in proniosome powders, which were were prepared with an equimolar ratio of span 60/cholesterol and four different carriers, namely, maltodextrin, mannitol, lactose, and pullulan.⁷⁴ The study showed that the niosomes produced by hydration and sonication of the proniosomes were small (≤200 nm) and quite homogeneously dispersed (PDI ≤0.3). The encapsulation efficiency of CTX in formulations varied between 55.3 \pm 1.8% and 74.1 \pm 2.7% after hydration and sonication. Although light and high temperatures affected the stability of CTX drastically, encapsulation in proniosomes retarded its degradation. This formulation can provide convenient, nontoxic, and inexpensive vehicles for dissolving and stabilizing CTX in functional food products.

O-padimate is a UV-B filter widely used as a sunscreen agent. A study investigated the combined influence of 3 independent variables in the preparation of *O*-padimate proniosomes, which were prepared by the slurry method with span: Brij, surfactant. The developed gels were characterized for vesicle size, morphology and EE, skin permeation studies, rheological properties, and sun protection factor (SPF). Results reveal that optimized *O*-padimate proniosomal formulations showed high SPF and low transepidermal water loss.⁷⁵

Rutin (Rut) is a natural flavonol that has various therapeutic properties including antioxidant and antitumor activities. Rut PNG for cutaneous applications was designed to improve the poor aqueous solubility of Rut. The gel was prepared by the coacervation phase-separation method and complies with the standard requirements in terms of PS (140.5 \pm 2.56 nm), ZP (-27.33 \pm 0.09 mV), encapsulation capacity (>50%), pH (7.002 \pm 0.18), and rheological properties. The results showed high biocompatibility of the gel on the 3D reconstructed human epidermis model characterized by increased viability of the cells and lack of irritant and phototoxic potential. The values on 2D cells confirmed the preferential cytotoxic effect of Rut on melanoma cells (IC₅₀ value: 8.601 μ M, nuclear fragmentation) compared with normal keratinocytes. Our data suggest that the

PNG is a promising drug carrier for Rut in the management and prevention of skin disorders.⁷⁶

Clinical trials with proniosomes

Table 2 Brief description

Selected proniosomal formulation of VP was tested in vivo to compare the pharmacokinetics of VP from a proniosomal patch containing VP (treatment A) to an oral commercial tablet containing the same dose of VP (treatment B) using a non-blind. two treatment, two-period, randomized, crossover design.77 Twelve healthy non-smoking male volunteers (26-37 years, 78-96 kg) participated in the study and were randomly assigned to one of the two treatment groups of equal size. The study results showed significant differences in the shape of the concentrationtime courses between the two treatments. For VP oral tablet a rapid sharp peak was found at 1.5 h followed by a fast decline in plasma drug levels. However, for the VP proniosomal patch, the absorption was much slower and extended over a longer period. Moreover, the patch exhibited higher drug levels in the plasma from 6 to 12 h compared to the tablet. The average C____ was significantly lower (p<0.05) for the patch (12.44 ± 1.87 ng/mL) compared with the oral tablet (63.69 ± 8.32 ng/mL), while t_{max} was significantly higher (p(0.05) in the case of the transdermal patch (12 h) compared with the oral tablet (1.5 h). The extent of absorption of VP from the patch expressed by AUC₀, was determined to be about 101% larger and statistically significantly different compared to the oral tablet. The relative bioavailability of VP proniosomal patch to the oral tablet was estimated to be on average 206%. The elimination half-lives of VP after oral and transdermal administration were 1.36 ± 0.27 h and 13.94 ± 1.2 h, respectively, and were statistically significant (p<0.01).77

TRT-loaded proniosomes were prepared by the slurry method with the help of span 60 and D-sorbitol, span 40, cholesterol
95% stabilized, and tween 20. The planned PNG was studied
clinically on 12 Egyptian patients aged more than 18 years [2
males and 10 females; with an average of 20 (\pm 4 years) with
acne (papules, closed comedones, and open comedones)]
on their face. ⁶⁴ The result showed only very slight erythema
(score: 0.143 \pm 0.377) for TRT PNG compared to 0.025% TRT
gel (score:1.70 ± 0.755). Similarly, the marketed product
displayed an erythema score of 1.50 \pm 0.534 and could not
diminish the irritation caused by topical application of TRT. The
overall improvement of the individual lesions was also better
than the marketed product during the 4 week study period.

Niosomal (hydrated form of proniosome) benzoyl peroxide (BPO) and clindamycin (CL) lotion was prepared and compared with niosomal CL in *acne vulgaris*. In both cases, the concentration of the drug was 1%. A double-blind clinical trial study on 100 patients with acne vulgaris was conducted in Afzalipour Hospital in Kerman (Iran).72 The efficacy of treatment protocols was evaluated in the 2nd, 4th, 8th, and 12th weeks of treatment by counting lesions (severity and grading acne lesions) and quality of life. Furthermore, the side effects were evaluated at each treatment visits. The reduction in the mean percentage of acne lesions in the case group (treated with BPO 1% and CL 1%) (64.21%) was higher than that the control group (treated with niosomal CL 1%) (59.04%), but the statistical difference was not significant. A sum of excellent and good results was found in 80% and 76.1% of the case and control groups, respectively (p= 0.377). Also, adding BPO to the treatment formulation in the case group did not increase adverse effects, as the statistical difference between the 2 groups was not significant (Table 3).78

Patent application IDs Inventors		Title	Patent description in brief	References
CN103340823A	Qiang L, Bin Z, Li L, Hongxia Z, Shasha L, Xiao J	Formulation of paeonol proniosomes and preparing method thereof	Paeonol proniosomes are an effective method for solving the problems of easy aggregation and easy fusion of niosomes and leakage of drugs in a solution state and a storage process of the non-ionic surfactant noisomes. The paeonol proniosomes can be directly applied to skins, and also can be prepared into a gel agent, an ointment, a patch and other transdermal drug delivery formulations	79
1288/DEL/2012	Yadav K, Yadav D, Nanda S, Saroha K	Curcumin proniosomal/ niosomal formulation, method for it is preparation and use thereof	Niosomal/proniosomal vesicular system can improve the dispersibility of curcumin and thus providing advantage to make curcumin in administrable or applicable form for it is cosmetic/therapeutic/medical purposes	80
3228/DEL/2012	Garg M, Joon M	Novel proniosomal gel of Withania somnifera	This invention relates to preparation of proniosomal gel of standardised <i>Withania somnifera</i> leaves extract, for better bioavailability, anti-inflammatory activity, to be used as transdermal application	81

Patent application IDs	Inventors	Title	Patent description in brief	References
3231/DEL/2012	Garg M, Garg P	Novel ursolic acid loaded proniosomal gel and method of preparation thereof	Ursolic acid is a well-known anti- inflammatory bioactive compound. This invention relates to the preparation of ursolic acid loaded proniosomal gel by coacervation phase separation method	82
U.S. Patent US 6051250A	Ribier A, Jean- Thierry S	Process for the stabilization of vesicles of amphiphilic lipid(s) and composition for topical application containing the said stabilized vesicles	This invention relates to a process for stabilization of vesicles formed from a lipid-phase membrane containing at least one ionic and/or non-ionic amphiphilic lipid encapsulating an aqueous phase, in the form of a dispersion in an aqueous phase, by addition of at least one stabilizing agent to the aqueous dispersion phase	83
U.S. Patent US 4830857A, 1989	Handjani R, Ribier A, Vanlerberghe G, Zabotto S, Griat J	Cosmetic and pharmaceutical compositions containing niosomes and a water-soluble polyamide, and a process for preparing these compositions	This invention relates to a composition consisting of a dispersion in an aqueous medium of niosome and/ or liposome spherules, within which an aqueous phase is encapsulated, at least one portion of the spherules being niosomes. The formulation is usable in cosmetics or pharmaceuticals	84
CN105311638A	Jianfang F, Kaili H, Helong L, Mei L, Luting W	Drug-carrying precursor vesicle, preparation method and application thereof	In this invention drug-carrying precursor vesicle can form homogeneous and mono-dispersed vesicles after hydration, has excellent stability, can significantly promote oral absorption and improve the bioavailability of poorly soluble drugs, and also has significant slow-releasing effect and is suitable for large-scale production	85
WO2000042987A8	Singh A and Jain R	Targeted vesicular constructs for cytoprotection and treatment of <i>Helicobacter</i> <i>pylori</i> infections	A novel composition for targeted vesicular constructs for treatment of <i>H.</i> <i>pylori</i> infections and for protection of the cell is disclosed	86
KR20040058196A	Young C, Lee W, Keith, Kwang-Ho	Pro-micelle pharmaceutical compositions	The present invention provides a composition containing a pharmaceutically active drug encapsulated with an esterified C_1-C_{18} fatty acid membrane. The result of the conversion of the micelles (pro- micelle) - are pharmaceutically that effectively deliver the active drug in systemic circulation procedure	87
EG24388A	Abd-Elbary A, El- laithy HM, Tadros MI	A method for the preparation of nebulizable micronized niosomes of cromolyn sodium using non-ionic surfactants	This invention involves the controlling the rate of drug release after inhalation so that biological half-life could be increased, consequently number of doses (4 - 6 daily) should be reduced	88

CONCLUSION

Proniosomes could be a promising vesicular drug delivery method for the future. They are one of the drug carriers in vesicular drug delivery methods, which is a better alternative to liposomal drug delivery due to its controlled and sustained action and provide better physical, chemical stability, and potentially scalable for commercial viability. They offer excellent potential for improved drug delivery through versatile routes such as oral, parenteral, dermal, transdermal, ocular, vaginal, pulmonary, and nasal by overcoming the permeation barriers faced by several drugs. Different types of unit dosages form like tablets, capsules, and beads can be prepared with the dry form of proniosomes. Due to the versatility of proniosomes, they are widely investigated as drug carriers. There is a lot of scope to investigate new carrier materials for the preparation of proniosomes and their potential remains to be investigated to the full extent.

Ethics

Peer-review: Externally and internally peer-reviewed.

Authorship Contributions

Concept: M.A., Design: M.A., F.A., Data Collection or Processing: F.A., Analysis or Interpretation: M.A., F.A., Literature Search: M.A., F.A., Writing: M.A., F.A.

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REFERENCES

- 1. Singla S, Harikumar S, Aggarwal G. Proniosomes for penetration enhancement in transdermal system. Int J Drug Dev Res. 2012;4:1-13
- Hu C, Rhodes DG. Proniosomes: a novel drug carrier preparation. Int J Pharm. 1999;185:23-35. Corrected and republished in: Int J Pharm. 2000;206:110-22.
- Naik A, Kalia YN, Guy RH. Transdermal drug delivery: overcoming the skin's barrier function. Pharm Sci Technol Today. 2000;3:318-326.
- Hiroyuki O, Makino K, eds. Colloid and interface science in pharmaceutical research and development. Elsevier, 2017.
- Prausnitz MR, Langer R. Transdermal drug delivery. Nat Biotechnol. 2008;26:1261-1268.
- Walve JR, Rane BR, Gujrathi NA, Bakaliwal SR, Pawar SP. Proniosomes: a surrogate carrier for improved transdermal drug delivery system. Int J Res Ayurveda Res. 2011;2:743-750.
- Sivaprasad SN, Kumar PL, Srinivas M, Brahmaiah B, Nama S. Proniosome: a novel approach to vesicular drug delivery system. Int J Drug Discov. 2013;3:85-90.
- 8. Nasseri B. Effect of cholesterol and temperature on the elastic properties of niosomal membranes. Int J Pharm. 2005;300:95-101.
- 9. Uchegbu IF, Vyas SP. Non-ionic surfactant-based vesicles (niosomes) in drug delivery. Int J Pharm. 1998;172:33-70.
- Bouwstra JA, van Hal DA, Hofland HEJ, Junginger HE. Preparation and characterization of nonionic surfactant vesicles. Colloids Surf A Physicochem Eng Asp. 1997;123-124:71-80.

- Pardakhty A, Varshosaz J, Rouholamini A. *In vitro* study of polyoxyethylene alkyl ether niosomes for delivery of insulin. Int J Pharm. 2007;328:130-141.
- Morakul B, Junyaprasert VB. Proniosomes: an effective carrier for dermal and transdermal delivery. Songklanakarin J Sci Technol. 2020;42:1171-1186.
- Bhardwaj P, Tripathi P, Gupta R, Pandey S. Niosomes: a review on niosomal research in the last decade. J Drug Deliv Sci Technol. 2020;56:Part A:101581.
- Ruckmani K, Sankar V. Formulation and optimization of zidovudine niosomes. AAPS PharmSciTech. 2010;11:1119-1127.
- Sahoo RK, Biswas N, Guha A, Kuotsu K. Maltodextrin based proniosomes of nateglinide: bioavailability assessment. Int J Biol Macromol. 2014;69:430-434.
- Akhilesh D, Faishal G, Kamath JV. Comparative study of carriers used in proniosomes. Int J Pharm Chem Sci. 2012;1:164-173.
- Ahmad MZ, Akhter S, Mohsin N, Abdel-Wahab BA, Ahmad J, Warsi MH, Rahman M, Mallick N, Ahmad FJ. Transformation of curcumin from food additive to multifunctional medicine: nanotechnology bridging the gap. Curr Drug Discov Technol. 2014;11:197-213.
- Blazek-Welsh AI, Rhodes DG. SEM imaging predicts quality of niosomes from maltodextrin-based proniosomes. Pharm Res. 2001;18:656-661.
- Mishra A, Kapoor A, Bhargava S. Proniosomal gel as a carrier for improved transdermal drug delivery. Asian J Pharm Life Sci. 2011;1:370-379.
- Jadhav KR, Pawar AY, Bachhav AA, Ahire SA. Proniosomes: a novel non-ionic pro-vesicles as potential drug carrier. Asian J Pharm Sci. 2016;10(Suppl):S210.
- Azeem A, Jain N, Iqbal Z, Ahmad FJ, Aqil M, Talegaonkar S. Feasibility of proniosomes-based transdermal delivery of frusemide: formulation optimization and pharmacotechnical evaluation. Pharm Dev Technol. 2008;13:155-163.
- Malhotra M, Jain NK. Niosomes as drug carriers. Indian Drugs. 1994;31:81-86.
- Biju SS, Talegaonkar S, Mishra PR, Khar RK. Vesicular systems: an overview. Indian J Pharm Sci. 2006;68:141-153.
- Keservani RK, Sharma AK, Ayaz MD. Novel drug delivery system for the vesicular delivery of drug by the niosomes. Int J Res Control Release. 2011;1:1-8.
- Kumar GP, Rao PR. Nonionic surfactant vesicular systems for effective drug delivery – an overview. Acta Pharm Sinica B. 2011; 1:208-219.
- Abd-Elbary A, El-Laithy HM, Tadros MI. Sucrose stearate-based proniosome-derived niosomes for the nebulisable delivery of cromolyn sodium. Int J Pharm. 2008;357:189-198.
- Vardhani S, Nirosha M, Chandrashekar KB. Proniosomal gel- an effective approach for topical and transdermal drug delivery. Int J Res Pharm Sci. 2016;7:179-183.
- 28. Radha GV, Rani TS, Sarvani B. A review on proniosomal drug delivery system for targeted drug action. J Basic Clin Pharm. 2013;4:42-48.
- 29. Yasam VR, Jakki SL, Natarajan J, Kuppusamy G. A review on novel vesicular drug delivery: proniosomes. Drug Deliv. 2014;21:243–249.
- Ammar HO, Ghorab M, El-Nahhas SA, Higazy IM. Proniosomes as a carrier system for transdermal delivery of tenoxicam. Int J Pharm. 2011;405:142-152.

- Song S, Tian B, Chen F, Zhang W, Pan Y, Zhang Q, Yang X, Pan W. Potentials of proniosomes for improving the oral bioavailability of poorly water-soluble drugs. Drug Dev Ind Pharm. 2015;41:51-62.
- Shehata TM, Abdallah MH, Ibrahim MM. Proniosomal oral tablets for controlled delivery and enhanced pharmacokinetic properties of acemetacin. AAPS PharmSciTech. 2015;16:375-383.
- Abdelbary GA, Aburahma MH. Oro-dental mucoadhesive proniosomal gel formulation loaded with lornoxicam for management of dental pain. J Liposome Res. 2015;25:107-121.
- Teaima MH, Yasser M, El-Nabarawi MA, Helal DA. Proniosomal telmisartan tablets: formulation, *in vitro* evaluation and *in vivo* comparative pharmacokinetic study in rabbits. Drug Des Devel Ther. 2020;14:1319-1331.
- Kuzmov A, Minko T. Nanotechnology approaches for inhalation treatment of lung diseases. J Control Release. 2015;219:500-518.
- Khalil RM, Abdelbary GA, Basha M, Awad GE, El-Hashemy HA. Design and evaluation of proniosomes as a carrier for ocular delivery of lomefloxacin HCl. J Liposome Res. 2017;27:118-129.
- Li Q, Li Z, Zeng W, Ge S, Lu H, Wu C, Ge L, Liang D, Xu Y. Proniosomederived niosomes for tacrolimus topical ocular delivery: *in vitro* cornea permeation, ocular irritation, and *in vivo* anti-allograft rejection. Eur J Pharm Sci. 2014;62:115-123.
- Aboali FA, Habib DA, Elbedaiwy HM, Farid RM. Curcumin-loaded proniosomal gel as a biofreindly alternative for treatment of ocular inflammation: *in-vitro* and *in-vivo* assessment. Int J Pharm. 2020;589:119835.
- Emad Eldeeb A, Salah S, Ghorab M. Proniosomal gel-derived niosomes: an approach to sustain and improve the ocular delivery of brimonidine tartrate; formulation, *in-vitro* characterization, and *in-vivo* pharmacodynamic study. Drug Deliv. 2019;26:509-521.
- Fouda NH, Abdelrehim RT, Hegazy DA, Habib BA. Sustained ocular delivery of dorzolamide-HCl via proniosomal gel formulation: *in-vitro* characterization, statistical optimization, and *in-vivo* pharmacodynamic evaluation in rabbits. Drug Deliv. 2018;25:1340-1349.
- Hao Y, Zhao F, Li N, Yang Y, Li K. Studies on a high encapsulation of colchicine by a niosome system. Int J Pharm. 2002;244:73-80.
- Elhissi A, Hidayat K, Phoenix DA, Mwesigwa E, Crean S, Ahmed W, Faheem A, Taylor KM. Air-jet and vibrating-mesh nebulization of niosomes generated using a particulate-based proniosome technology. Int J Pharm. 2013;444:193-199.
- Harrison PF. Microbicides: forging scientific and political alliances. AIDS Patient Care STDS. 2000;14:199-205.
- Radha GV, Vinisha V, Rajkumar J, Ghosh A. Design and evaluation of topical vaginal proniosomal formulations of tenofovir disoproxil fumarate for HIV prevention. Int J Pharm Res. 2019;10:94-102.
- Abdou EM, Ahmed NM. terconazole proniosomal gels: effect of different formulation factors, physicochemical and microbiological evaluation. J Pharm Drug Deliv Res. 2016;5:1.
- Verma P, Prajapati SK, Yadav R, Senyschyn D, Shea PR, Trevaskis NL. Single intravenous dose of novel flurbiprofen-loaded proniosome formulations provides prolonged systemic exposure and antiinflammatory effect. Mol Pharm. 2016;13:3688-3699.
- Khudair N, Agoun A, Elrayess MA, Najlah M, Younes HM, Elhissi
 A. Letrozole-loaded nonionic surfactant vesicles prepared via a

slurry-based proniosome technology: formulation development and characterization. J Drug Deliv Sci Technol. 2020;58:101721.

- Chandra A, Sharma PK. Proniosome based drug delivery system of piroxicam. Afr J Pharm. Pharmacol. 2008;2:184-190.
- Solanki AB, Parikh JR, Parikh RH. Preparation, optimization and characterization of ketoprofen proniosomes for transdermal delivery. Int J Pharm Sci. Nanotech. 2009;2:413-420.
- Mahrous GM. Proniosomes as a drug carrier for transdermal delivery of meloxicam. Bull Pharm Sci. 2010;33:131-140.
- Alam MI, Baboota S, Kohli K, Ali J, Ahuja A. Pharmacodynamic evaluation of proniosomal transdermal therapeutic gel containing celecoxib. ScienceAsia. 2010;36:305-311.
- Abu El-Enin ASM, Khalifa MKA, Dawaba AM, Dawaba HM. Proniosomal gel-mediated topical delivery of fluconazole: development, *in vitro* characterization, and microbiological evaluation. J Adv Pharm Technol Res. 2019;10:20-26.
- Fang JY, Yu SY, Wu PC, Huang YB, Tsai YH. *In vitro* skin permeation of estradiol from various proniosome formulations. Int J Pharm. 2001;215:91-99.
- Mehta M, Dureja H, Garg M. Development and optimization of boswellic acid-loaded proniosomal gel. Drug Deliv. 2016;23:3072-3081.
- 55. Eltellawy YA, El-Kayal M, Abdel-Rahman RF, Salah S, Shaker DS. Optimization of transdermal atorvastatin calcium - loaded proniosomes: restoring lipid profile and alleviating hepatotoxicity in poloxamer 407-induced hyperlipidemia. Int J Pharm. 2021;593:120163.
- Nemr AA, El-Mahrouk GM, Badie HA. Development and evaluation of proniosomes to enhance the transdermal delivery of cilostazole and to ensure the safety of its application. Drug Dev Ind Pharm. 2021;47:403-415.
- Tareen FK, Shah KU, Ahmad N, Ur Rehman A, Shah SU, Ullah N. Proniosomes as a carrier system for transdermal delivery of clozapine. Drug Dev Ind Pharm. 2020:1-24.
- Sarfaraz M, Goel T, Doddayya H. Formulation and evaluation of galantamine hydrobromide proniosome gel for Alzheimer's disease. JDDTAO. 2020;10(Suppl):68-74.
- Khatoon M, Sohail MF, Shahnaz G, Ur Rehman F, Fakhar-Ud-Din, Ur Rehman A, Ullah N, Amin U, Khan GM, Shah KU. Development and evaluation of optimized thiolated chitosan proniosomal gel containing duloxetine for intranasal delivery. AAPS PharmSciTech. 2019;20:288.
- Farris PK. Cosmeceuticals and cosmetic practice. Oxford, UK: Wiley-Blackwell; 2014.
- Souto EB, Müller RH. Cosmetic features and applications of lipid nanoparticles (SLN, NLC). Int J Cosmet Sci. 2008;30:157-165.
- Blazek-Welsh AI, Rhodes DG. Maltodextrin-based proniosomes. AAPS PharmSci. 2001;3:E1.
- Manosroi J, Chankhampan C, Kitdamrongtham W, Zhang J, Abe M, Akihisa T, Manosroi W. *In vivo* anti-ageing activity of cream containing niosomes loaded with purple glutinous rice (*Oryza sativa* Linn.) extract. Int J Cosmet Sci. 2020;42:622-631.
- Rahman SA, Abdelmalak NS, Badawi A, Elbayoumy T, Sabry N, El Ramly A. Formulation of tretinoin-loaded topical proniosomes for treatment of acne: *in-vitro* characterization, skin irritation test and comparative clinical study. Drug Deliv. 2015;22:731-739.

- Chainani-Wu N. Safety and anti-inflammatory activity of curcumin: a component of tumeric (*Curcuma longa*). J Altern Complement Med. 2003;9:161-168.
- Shehata TM, Ibrahim MM, Elsewedy HS. Curcumin niosomes prepared from proniosomal gels: *in vitro* skin permeability, kinetic and *in vivo* studies. Polymers (Basel). 2021;13:791.
- Yadav NK, Nanda S, Sharma G, Katare OP. Systematically optimized coenzyme q10-loaded novel proniosomal formulation for treatment of photo-induced aging in mice: characterization, biocompatibility studies, biochemical estimations and anti-aging evaluation. J Drug Target. 2016;24:257-271.
- Schlich M, Lai F, Pireddu R, Pini E, Ailuno G, Fadda AM, Valenti D, Sinico C. Resveratrol proniosomes as a convenient nanoingredient for functional food. Food Chem. 2020;310:125950.
- de Sá Coutinho D, Pacheco MT, Frozza RL, Bernardi A. Anti-inflammatory effects of resveratrol: mechanistic insights. Int J Mol Sci. 2018;19:1812.
- Salehi B, Mishra AP, Nigam M, Sener B, Kilic M, Sharifi-Rad M, Fokou PVT, Martins N, Sharifi-Rad J. Resveratrol: a double-edged sword in health benefits. Biomedicines. 2018;6:91.
- Smoliga JM, Baur JA, Hausenblas HA. Resveratrol and health-a comprehensive review of human clinical trials. Mol Nutr Food Res. 2011;55:1129-1141.
- Zupančič Š, Lavrič Z, Kristl J. Stability and solubility of trans-resveratrol are strongly influenced by pH and temperature. Eur J Pharm Biopharm. 2015;93:196-204.
- Tanaka T, Shnimizu M, Moriwaki H. Cancer chemoprevention by carotenoids. Molecules. 2012;17:3202-3242.
- Ravaghi M, Sinico C, Razavi SH, Mousavi SM, Pini E, Fadda AM. Proniosomal powders of natural canthaxanthin: preparation and characterization. Food Chem. 2017;220:233-241.
- 75. Sandeep G, Raju J, Subba Rao D, Vamshi Krishna M. Proniosomal sunscreen gel-based formulation: a promising approach for improving quality of life (QOL). Reconstructive Surgery and Anaplastology. Longdom Publishing SL; -1 [cited 2021May19]. Available from: https:// www.longdom.org/proceedings/proniosomal-sunscreen-gel-basedformulation--a-promising-approach-for-improving-quality-of-lifeqol-613.html

- 76. Pinzaru I, Tanase A, Enatescu V, Coricovac D, Bociort F, Marcovici I, Watz C, Vlaia L, Soica C, Dehelean C. Proniosomal gel for topical delivery of rutin: preparation, physicochemical characterization and *in vitro* toxicological profile using 3D reconstructed human epidermis tissue and 2D cells. Antioxidants (Basel). 2021;10:85.
- El-Laithy HM, Shoukry O, Mahran LG. Novel sugar esters proniosomes for transdermal delivery of vinpocetine: preclinical and clinical studies. Eur J Pharm Biopharm. 2011;77:43-55.
- Mohammadi S, Pardakhty A, Khalili M, Fathi R, Rezaeizadeh M, Farajzadeh S, Mohebbi A, Aflatoonian M. Niosomal benzoyl peroxide and clindamycin lotion *versus* niosomal clindamycin lotion in treatment of acne vulgaris: a randomized clinical trial. Adv Pharm Bull. 2019;9:578-583.
- 79. Qiang L, Bin Z, Li L, Hongxia Z, Shasha L, Xiao J. Formulation of paeonol proniosomes and preparing method thereof. CN103340823A.
- Yadav K, Yadav D, Nanda S, Saroha K. Curcumin proniosomal/niosomal formulation, method for its preparation and use thereof. Indian Patent. 1288/DEL/2012.
- Garg M, Joon M. Novel Proniosomal gel of Withania somnifera. Indian Patent. 3228/DEL/2012.
- Garg M, Garg P. Novel ursolic acid loaded proniosomal gel and method of preparation thereof. Indian Patent. 3231/DEL/2012.
- Ribier A, Jean-Thierry S. Process for the stabilization of vesicles of amphiphilic lipid(s) and composition for topical application containing the said stabilized vesicles. U.S. Patent. US 6051250A
- Handjani R, Ribier A, Vanlerberghe G, Zabotto S, Griat J. Cosmetic and pharmaceutical compositions containing niosomes and a water soluble polyamide, and a process for preparing these compositions. U.S. Patent. US 4830857A.
- Jianfang F, Kaili H, Helong L, Mei L, Luting W. Drug-carrying precursor vesicle, preparation method and application thereof. CN105311638A.
- Singh A, Jain R. Targeted vesicular constructs for cytoprotection and treatment of *H. pylori* infections. WO2000042987A8.
- Young C, Lee W, Keith, Kwang-Ho. Pro-micelle pharmaceutical compositions. KR20040058196A.
- Abd-Elbary A, El-laithy HM, Tadros MI. A method for the preparation of nebulizable micronized niosomes of cromolyn sodium using nonionic surfactants. EG24388A.